M.Sc.

in

Microbiology

COURSE STRUCTURE &SYLLABI

(Academic Session 2021-22 onwards)

Department of Life Science
School of Basics Sciences and Research
SHARDA UNIVERSITY

SUMMARY SHEET

Teaching Department	**:**	Life Science
School	**:**	School of Basic Sciences and Research
Name of Course	**:**	M.Sc. in Microbiology
Duration	**:**	Two Years
Total number of Credits	**:**	90

Term I

S. No.	Subject Code	Subjects	Teaching Load			Credits
			L	T	P	
THEORY SUBJECTS						
1	MMB101	Microbial Diversity	4	0	0	4
2	MMB102	Molecular Biology	4	0	0	4
3	MMB103	Microbial Metabolism	4	0	0	4
4	MMB104	Enzymology	4	0	0	4
5	MSB124	IPR	4	0	0	4
PRACTICALS						
1	MMB153	Microbial Diversity Lab	0	0	3	2
2	MMB159	Enzymology Lab	0	0	3	2
3	MMB157	Molecular Biology lab	0	0	3	2
TOTAL						**26**

Term II

S. No.	Subject Code	Subjects	Teaching Load			Credits
			L	T	P	
THEORY SUBJECTS						
1	MSB116	Bio-instruments	4	0	0	4
2	MMB110	Mycology, Phycology and Virology	4	0	0	4
3	MMB108	Recombinant-DNA Technology	4	0	0	4
4	MMB107	Bacteriology	4	0	0	4
5	MSB125	Bioinformatics	4	0	0	4
PRACTICALS						
1	MSB160	Bioinstrumentation Lab	0	0	3	2
2	MMB156	RDT Lab	0	0	3	2
3	MMB160	Mycology, Phycology and Virology Lab	0	0	3	2
TOTAL						**26**

Term III

S. No.	Subject Code	Subjects	Teaching Load			Credits
			L	T	P	
THEORY SUBJECTS						
1	MMB201	Environmental Microbiology & Waste Management	4	0	0	4
2	MMB202	Infection, Immunity and Diagnostics	4	0	0	4
3	MSB207	Microbial Biotechnology	4	0	0	4
4	MMB207	Fermentation and downstream processes	4	0	0	4
5	MMB208	Food Microbiology	**2**	**0**	**0**	**2**
PRACTICALS						
1	MSB259	Microbial Biotechnology Lab	0	0	3	2
2	MMB255	Immunology lab	0	0	3	2
3	MMB260	Fermentation Technology Lab	0	0	3	2
4	CCU401	Community Connect	0	0	2	2
TOTAL						**26**

Term IV

S. No.	Subject Code	Subjects	Teaching Load			Credits
			L	T	P	
1	MSB261	Dissertation / Project work / Industrial Training	0	0	18	12

MMB101: Microbial Diversity

L-T-P: 4-0-0 Credit – 4

School: SBSR		**Batch: 2020 – 22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 01**	
1	Course Code	**MMB101**	
2	Course Title	**Microbial Diversity**	
3	Credits	4	
4	Contact Hrs (L-T-P)	4-0-0	
	Course Status	**Compulsory**/Elective/Open Elective	
5	Course Objective	1. Diversity of Microbial World 2. Classification system of microorganisms 3. General characteristic features of archaea, eubacteria, algae and fungi 4. Mode of reproduction of eubacteria, algae and fungi	
6	Course Outcomes	After studying this course, students will be able to CO1: Determine general characteristics of acellular and cellular microorganisms, classification system as well as difference between prokaryotes and eukaryotes CO2: Summarize the diversity, characteristic features and significance of archaea CO3: Describe the diversity, characteristic features and significance of eubacteria CO4: Determine the general characteristics, cellular structure as well as potential applications of algae CO5: Analyze the general characteristics, mode of reproduction as well as mode of reproduction in fungi CO6: Compare the characteristic features, mode of reproduction and significance of various microbial living systems	
7	Course Description	The course comprises of general and characteristic features of diverse microbial living systems such as acellular and cellular microbes, archaebacteria, eubacteria, algae and fungi.	
8	Outline syllabus		CO Mapping
	Unit 1	**Diversity of Microbial World and Microbial Classification**	
	a	General characteristics of different groups: Acellular microorganisms (Viruses, Viroids, Prions) and cellular microorganisms (Bacteria, Algae, Fungi and Protozoa)	CO1, CO6
	b	Systems of classification. Binomial Nomenclature, Whittaker's five kingdom and Carl Woese's three kingdom classification systems and their utility	CO1, CO6

	c	Difference between prokaryotic and eukaryotic microorganisms			CO1, CO6
	Unit 2	**Archaea**			
	a	Occurrence, diversity			CO2, CO6
	b	Characteristic features, significance			CO2, CO6
	c	Potential applications of different groups of archaebacteria (e.g. methane generation, ultrafiltration membranes, desulphurization of coal and crude oil, bioleaching of metals, enzymes, compatible solutes and others)			CO2, CO6
	Unit 3	**Bacteria**			
	a	Occurrence, diversity, characteristic features			CO3, CO6
	b	Significance and potential applications of various groups of bacteria			CO3, CO6
	c	Very precise account of typical eubacteria			CO3, CO6
	Unit 4	**Algae**			
	a	General characteristics of algae including occurrence, thallus organization			CO4, CO6
	b	algae cell ultra-structure, pigments, flagella, eyespot food reserves and vegetative, asexual and sexual reproduction			CO4, CO6
	c	potential applications (e.g. Importance of algae in production of algal pigments, biofuels, hydrogen production, important bioactive molecules, role of algae in sustainable environment)			CO4, CO6
	Unit 5	**Fungi**			
	a	General characteristics of fungi including habitat, distribution, nutritional requirements,			CO5, CO6
	b	fungal cell ultra- structure, thallus organization and aggregation, fungal wall structure and synthesis			CO5, CO6
	c	asexual reproduction and sexual reproduction. Potential applications of different groups of fungi			CO5, CO6
	Mode of examination	**Theory**			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Textbook/s*	1. Pelczar MJ, Chan ECS and Krieg NR. (1993). Microbiology. 5th edition. McGraw Hill Book Company.			
	Other References	1. Atlas RM. (1997). Principles of Microbiology. 2nd edition. WM.T. Brown Publishers. 2. Kumar HD. (1990). Introductory Phycology. 2nd edition. Affiliated East Western Press. 3. Alexopoulos CJ, Mims CW, and Blackwell M. (1996). Introductory Mycology. 4th edition. John and Sons, Inc.			

MMB102: Molecular Biology

L-T-P: 4-0-0 Credit: 4

School: SBSR		**Batch: 2020 – 22**
Program: M.Sc.		**Current Academic Year: 2020-21**
Branch: Microbiology		**Semester: 01**
1	Course Code	**MMB102**
2	Course Title	**MOLECULAR BIOLOGY**
3	Credits	4
4	Contact Hours (L-T-P)	4-0-0
	Course Status	**Compulsory** /Elective/Open Elective
5	Course Objective	1. Understand DNA as genetic information carrier, its evolution, structure, synthesis and packaging. 2. Describe various mechanisms involved in gene expression at transcriptional and translational levels. 3. Observe different perspectives of gene regulation for therapeutic applications.
6	Course Outcomes	CO1: Understand DNA as genetic information carrier, its evolution, structure, synthesis and packaging. CO2: Examine RNA structure, its types and significance of the mechanisms involved in its complete synthesis. CO3: Describe key players in regulation of gene expression and post transcriptional modifications. CO4: Elaborate protein synthesis, post translational modifications and protein trafficking. CO5: Identify the roles of oncogenes and tumour suppressor genes in cancer development and thus finding the therapeutic molecular mechanisms for cancer treatment. CO6: Observe different perspectives of gene regulation in life processes at molecular level inside cell.
7	Course Description	This course will cover the major topics in Molecular Biology, including "DNA as genetic information carrier, its evolution, structure, synthesis and packaging", "RNA structure, its types and significance of the mechanisms involved in its complete synthesis", "key players in regulation of gene expression and post transcriptional modifications", "Elaborate protein synthesis, post translational modifications and protein trafficking", "Identify the roles of oncogenes and tumour suppressor genes in cancer development and thus finding the therapeutic molecular mechanisms for cancer treatment".
8	Outline syllabus	CO Mapping

	Unit 1	**Nucleic Acids as Genetic Information Carrier**			CO1, CO6
	A	Experimental evidence. DNA structure: historical aspects and current concepts			
	B	Melting of DNA, Replication: general principles, various modes of replication, isolation and properties of DNA polymerases, proof reading, continuous and discontinuous synthesis, Asymmetric & dimeric nature of DNA polymerases, synthesis of leading and lagging stands			
	C	Superhelicity in DNA, linking number, topological properties, mechanism of action of topoisomerases			
	Unit 2	**Transcription**			CO2, CO6
	A	General principles, basic apparatus, types of RNA polymerases, steps: initiation, elongation and termination			
	B	Structural features of RNA (rRNA, tRNA and mRNA) and relation to function. Peptidyl transferase activity of 23S tRNA. Polycistronic and monocistronic RNAs			
	C	Control of transcription by interaction between RNA polymerases and parameter regions, use of alternate sigma factors, controlled termination: attenuation and ant-termination			
	Unit 3	**Regulation of Gene Expression**			CO3, CO6
	A	Operon concept, catabolite repression instability of bacterial RNA, positive and negative regulation, inducers and co-repressors			
	B	Negative regulation - *E. coli* lac operon, positive regulation. *E. coli* ara operon; his and trp operons			
	C	Maturation and processing of RNA, methylation, cutting and trimming of rRNA; capping, polyadenylation and splicing of m RNA; cutting and modification of tRNA degradation system. Catalytic RNA group I and group II, intron splicing RNase P			
	Unit 4	**Translation**			CO4, CO6
	A	Prokaryotic and eukaryotic translation, mechanisms of initiation, elongation and termination, regulation of translation			
	B	Post-translational modifications of proteins			
	C	Protein localization, synthesis of secretory and membrane proteins, intracellular transportation of different proteins			
	Unit 5	**Oncogenes and Tumor Suppressor Genes**			CO5, CO6
	A	Homologous recombination, Holiday junction			
	B	DNA repair mechanisms			
	C	Oncogenes and Tumor suppressor genes- Viral and cellular oncogenes, tumor suppressor genes, structure, function and mechanism of tumor suppressor proteins; Role of p53 and other proteins in cancer, carcinogens and other transforming agents			
	Mode of examination	**Theory**/Jury/Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	

	Textbook/s*	Molecular biology of the Gene (4th Edition),J .D. Watson, N. H. Hopkins, J. W. Roberts,J.A. Steitz and A.M.	

MMB103: Microbial Metabolism

L-T-P: 4-0-0 Credit - 4

School: SBSR		**Batch: 2020 – 22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 01**	
1	Course Code	**MMB103**	
2	Course Title	**Microbial Metabolism**	
3	Credits	4	
4	Contact Hrs (L-T-P)	4-0-0	
	Course Status	**Compulsory** /Elective/Open Elective	
5	Course Objective	1. Metabolic pathways in microorganisms, study of bioenergetics, nature and significance of central metabolic pathways and also their regulation. 2. Central metabolic pathways as the backbone of other metabolic events in the cell, such as metabolism of nucleotide, lipids and protein. 3. Integration of all metabolic pathways. 4. Photosynthetic fixation of carbon and assimilation of some of the vital inorganic metals such as phosphorous, sulphur and nitrogen.	
6	Course Outcomes	After studying this course, students will be able to CO1: Determine standard free energy, hydrolysis of ATP and its role. Further the significance of metabolic regulation CO2: Evaluate metabolism of carbohydrates by different pathways CO3: Interpret the structure, functions and metabolism of different types of lipids CO4: Differentiate between de novo and salvage pathways for biosynthesis of purines and pyrimidines CO5: Determine photosynthetic fixation of carbon. CO6: Analyze and study various metabolic pathways in microorganisms	
7	Course Description	This course contains various metabolic pathways inside a microorganism such as metabolism of carbohydrates, lipids, nucleic acids and also carbon dioxide fixation. After studying course, students will be able to learn various metabolic processes going inside the body of microbes.	
8	Outline syllabus		CO Mapping
	Unit 1	**Energy, Enzymes and Regulation**	
	a	Energy and work, Laws of thermodynamics, Free energy and reactions	CO1, CO6
	b	Role of ATP in metabolism, Oxidation-reduction reactions and electron carrier	CO1, CO6
	c	Nature and significance of metabolic regulation, Metabolic channelling	CO1, CO6

	Unit 2	**Carbohydrate Metabolism**			
	a	Carbohydrates: Central pathways of metabolism - regulatory mechanisms, Bioenergetics and significance - EMP and alternate pathways: Entner-Doudoroff			CO2, CO6
	b	HMP and oxidative pentose phosphate, TCA cycle, Glyoxylate cycle			CO2, CO6
	c	Utilization of sugars and polysaccharides, Gluconeogenesis from TCA intermediates / amino acids / acetyl-CoA; Electron Transport Chain			CO2, CO6
	Unit 3	**Lipid Metabolism**			
	a	Lipids: fatty acids - structure, properties; classification of lipids, structure, properties, lipid composition of microorganisms			CO3, CO6
	b	Catabolism: Bioenergetics of β-oxidation of fatty acids, long chain fatty acids			CO3, CO6
	c	Anabolism: Biosynthesis of fatty acids: saturated, unsaturated, Biosynthesis of triglycerides, phospholipids, sterols			CO3, CO6
	Unit 4	**Nucleotide Biosynthesis**			
	a	Synthesis of purines, pyrimidines and nucleotides			CO4, CO6
	b	Purine biosynthesis, Pyrimidine biosynthesis			CO4, CO6
	c	Biosynthesis of nucleotide coenzymes			CO4, CO6
	Unit 5	**Use of Energy in Biosynthesis**			
	a	Photosynthetic fixation of CO_2, Carboxylation phase, Reduction phase, Regeneration phase			CO5, CO6
	b	Synthesis of sugars and polysaccharides, Assimilation of inorganic phosphorus, sulphur and nitrogen			CO5, CO6
	c	Nitrogen fixation, Synthesis of amino acids, Anaplerotic reactions			CO5, CO6
	Mode of examination	Theory			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Textbook/s*	Nelson D.L., Cox M. M., “Principles of Biochemistry” W. H. Freeman, 2012.			
	Other References	Stryer L., “Biochemistry”, W. H. Freeman, 2010. Jain JL., “Principles of Biochemistry”, S. Chand Publications.			

MMB104: Enzymology

L-T-P: 4-0-0 Credit – 4

School: SBSR		**Batch: 2020 – 22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 1**	
1	Course Code	**MMB 104**	
2	Course Title	**Enzymology**	
3	Credits	4	
4	Contact Hours (L-T-P)	4-0-0	
	Course Status	Compulsory	
5	Course Objective	With this Course the students 1. will acquire knowledge fundamental Knowledge of Enzymes 2. Will get useful exploitation of enzymes physical and kinetic properties 3. Use Enzymes biocatalysts in the biotransformation 4. Know the Industrial, Research and Therapeutic applications of Enzymes.	
6	Course Outcomes	After successfully completion of this course students will be able to: CO1: Define and Classify Enzymes and its fundamentals properties CO2: Examine Enzyme Kinetics, Perform and calculate enzyme specificity and activity CO3: Evaluate Enzyme Inhibition and its types, Competitive and Non-competitive inhibition and its significance CO4: Understand Allosteric Enzymes regulation, Covalent modification, Determine the role of co-enzymes, Enzyme constitution and immobilization CO5: Evaluate Applications of Enzymes in industry, Enzymes in clinical diagnostics. sensors for clinical processes and environmental, Microbial analyses, Engineered Enzymes. CO6: To analyse Enzymes principles, properties, Kinetics, Inhibition, Allosterism, Co-Enzymes, Engineered Enzymes, Application of Enzymes in various industries, research and therapeutic aspects	
7	Course Description	This course covers fundamentals to applications necessary for the useful exploitation of enzymes both as tools for the enzymatic analyses and as biocatalysts in the biotransformations on the unique structural-functional properties of enzymes and its microbial industrial and research utilization.	
8	Outline syllabus		CO Mapping
	Unit 1	**Properties of Enzymes**	CO1,6
	A	Classification of enzymes, Structural conformations of enzyme proteins	CO1,6
	B	Enzymes as biocatalysts, Catalytic power, Activation energy	CO1,6

	C	Substrate specificity, Mechanisms of enzyme action, Ribozymes and abzymes.			CO1,6
	Unit 2	**Enzyme Kinetics**			CO2,6
	A	Factors affecting rates of enzymatic reactions (pH, temperature, substrate concentration, enzyme concentration and reaction time)			CO2,6
	B	Overview of Michaelis-Menten equation and its transformation, Lineweaver-Burke plot			CO2,6
	C	Evaluation of kinetic parameters (K_M, V_{max}).			CO2,6
	Unit 3	**Enzyme Inhibition**			CO3,6
	A	Irreversible and reversible inhibition			CO3,6
	B	Competitive, non-competitive and un-competitive inhibition			CO3,6
	C	Enzyme inhibition kinetic studies, Determination of k_{cat}.			CO3,6
	Unit 4	**Regulation of Enzyme Activity**			CO4,6
	A	Allosterism, Kinetic analysis of allosteric enzymes			CO4,6
	B	Covalent modification, Feed-back inhibition, Membrane bound enzymes			CO4,6
	C	Isoenzymes and marker enzymes, Constitutive and inducible enzymes.			CO4,6
	Unit 5	**Applications of Microbial Enzymes**			CO5,6
	A	Microbial enzymes in textile, leather, wood industries and detergents			CO5,6
	B	Enzymes in clinical diagnostics and Enzyme sensors for clinical processes and environmental analyses			CO5,6
	C	Engineered enzymes, Enzymes as therapeutic agents.			CO5,6
	Mode of examination	Theory			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Textbook/s*	Palmer T., Bonner P. L., "Enzymes: Biochemistry, Biotechnology, Clinical Chemistry", Woodhead Publishing, 2007.			
	Other References	1. Copeland R. A., "Enzymes: A Practical Introduction to Structure, Mechanism, and Data Analysis", Wiley, 2006. 2. Guisán J. M., "Immobilization of Enzymes and Cells (Methods in Biotechnology)", Humana Press, 2010.			

MSB124: IPR (Intellectual Property Rights)

L-T-P: 4-0-0 Credit - 4

School : SBSR		**Batch : 2020– 22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 1**	
1	Course Code	MSB124	
2	Course Title	**Intellectual Property Rights**	
3	Credits	4	
4	Contact Hours (L-T-P)	4-0-0	
	Course Status	Compulsory	
5	Course Objective	To elucidate the ways of protection of intellectual property and research with the help of WIPO and its different treaties. To correlate different instruments of IP protection and their enforcement in different countries. To understand different quality management issues related to biotechnology	
6	Course Outcomes	By the end of this course students will be able to: CO1: Administer and follow the guidelines of WIPO. CO2: Understand the patents, copyrights and trademarks. CO3: Understand the character merchandising and franchising. CO4: Understand the utility of IPRs in biotechnology.	
7	Course Description	*Intellectual property* (IP) includes intangible creations of the human intellect, and primarily encompasses copyrights, patents, and trademarks. It also includes other types of rights, such as trade secrets, publicity rights, moral rights, and rights against unfair competition. Present paper deals with knowledge of types and protection of different IPRs.	
8	Outline syllabus		CO Mapping
	Unit 1	**Introduction to Intellectual Property Rights**	**CO1, CO4**
	A	The concept of intellectual property, Importance of IPR in biotechnology	
	B	WIPO- history, mission and activities, structure, administration.	
	C	Major International Instruments relating to the protection of IP; Berne Convention; Paris Convention; TRIPS	
	Unit 2	**Patents**	
	A	Patents-basic concepts; Non patentable inventions	CO2, CO3, CO4
	B	Procedure for registration , Term of patent , Rights of patentee	
	C	Patent Infringement and its remedy; Compulsory licenses and Government use of patent	

	Unit 3	**Copyrights**			**CO2, CO3, CO4,**
	A	Copyright and related rights;			
	B	Copyright piracy and infringement; Remedies of copyright piracy and infringement			
	C	Copyright Issues in Digital Environment			
	Unit 4	**Trademarks**			**CO2, CO3, CO4,**
	A	Definitions, Signs which serve as trademarks,			
	B	Trademark piracy, and counterfeiting; Character Merchandising.			
	C	Geographical Indication; Difference between GI and Trade Marks			
	Unit 5	**IPR in industries**			**CO3, CO4,**
	A	IPR strategies by different industries; E-Commerce and IPR issues			
	B	Case studies of Major IPR conflicts: Zara Vs Zara fashions; Yahoo Vs Yahoo India.			
	C	Case studies of Major IPR conflicts: AMUL Vs IMUL; Paytm Vs PayPal			
	Mode of examination	Theory			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Text book/s*	1. Managing intellectual capital: organizational, strategic and policy dimensions Oxford Univ. press 2005 Teece, David J.			
	Other References	2. Techniques used in Bio product analysis, Butterworth Heinemann Ltd, 2017. 3. Law relating to patents, trademarks, copyright designs geographical indications. Universal Law Publishing house by Wadehra, B.L.			

MMB153: Microbial Diversity Lab

L-T-P: 0-0-3 Credit - 2

School: SBSR		**Batch: 2020 – 22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 01**	
1	Course Code	**MMB153**	
2	Course Title	**Microbial Diversity Lab**	
3	Credits	2	
4	Contact Hours (L-T-P)	0-0-3	
	Course Status	**Compulsory/Elective**	
5	Course Objective	To learn methods of cell isolation from tissues and determine enzyme activity and inhibition of different proteins.	
6	Course Outcomes	CO1: Perform detection of protein from the given samples. CO2: Carry out an experiment for the detection of starch. CO3: Distinguish glucose from the given sample with the help of designed experiment. CO4: Design and conduct the experiment. CO5: Protein separation by chromatographic techniques. CO6: Plan and carry out the experiment. CO7: Carry out an experiment for the visualization of DNA on agarose gel. CO8: Design and conduct the experiment. CO9: Plan and carry out an experiment for the separation and quantification of fat from milk	
7	Course Description	To Plan and carry out the experiment and to learn methods of cell isolation from tissues and determine enzyme activity and inhibition of different proteins. Design and conduct the experiment.	
8	Outline syllabus		CO Mapping
	Unit 1	Isolation of individual cells from mixed culture	CO4,CO6,CO8
		Characterization based on shape and size of microbial colonies	
	Unit 2	Gram Stain Technique	CO1,CO8,CO6
		Differential and Cytological Staining	CO4,CO5,CO6
	Unit 3	Acid Fast Staining	CO2,CO5,CO6
		Catalase Test	CO1,CO5,CO6
	Unit 4	Carbohydrate Fermentation Test	CO4,CO5,CO6
		Bacterial Growth Curve	
	Unit 5	Methylene Blue Reductase Test	CO7,CO5,CO6

		Urease Test			CO7,CO6
	Mode of exam	Jury/Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		60%	0%	40%	
	Textbook/s*	Practical manual of Biotechnology by Ritu Mahajan, Jitendar Sharma, RK Mahajan, Vayu Education of India			
	Other References	Practical Microbiology by DK Maheshwari, S Chand Publications.			

MMB157: Molecular Biology Lab

L-T-P: 0-0-3 Credit - 2

School: SBSR		**Batch: 2020 – 22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester - 01**	
1	Course Code	MMB152	
2	Course Title	Molecular Biology Lab	
3	Credits	2	
4	Contact Hours (L-T-P)	0-0-3	
	Course Status	Compulsory	
5	Course Objective	1. To familiarize students with sterilization techniques and solution/media preparations etc. 2. To motivate students towards molecular techniques for better genome understanding. 3. To acquaint with principles, technical requirement, scientific and commercial applications in molecular biology. 4. Design and manage techniques for understanding interplay amongst macromolecules.	
6	Course Outcomes	CO1: Demonstrate safe laboratory practices and handle the equipment safely. CO2: Estimate the quality and quantity of nucleic acids. CO3: Amalgamation of tools for plasmid vectors and DNA uptake. CO4: Perform *in silico* analysis for studying genome. CO5: To design primers and carry out amplification of DNA by PCR.	
7	Course Description	The aim of this course is to acquaint the students about the versatile tools and techniques employed in molecular biotechnology. The course will also provide students with a hands-on understanding of how modern DNA-sequencing technology, along with bioinformatics tools, can be used to discover genetic differences and understand molecular function.	
8	Outline syllabus		CO Mapping
	Unit 1	**Practical based on introduction to molecular biology lab**	**CO1**
	A	Good lab practices in molecular biology laboratory.	
	B & C	Preparation of standard solutions for molecular biology experiments	
	Unit 2	**Isolation of Nucleic acids and quantification**	**CO2**

	A	Isolation of DNA from bacteria			
	B	Isolation of RNA from bacteria			
	C	Gel electrophoresis			
	Unit 3	**Practical related to preparation of plasmids and transformations**			**CO3**
	A	Plasmid isolation			
	B	Preparation of competent cells			
	C	Transformation of plasmid into competent cells			
	Unit 4	**Practical related to in silico analysis of genome**			**CO4**
	A	Sequence similarity search with freely available tools			
	B	Construction of phylogenetic tree			
	C	Identification of motifs and domain in sequences			
	Unit 5	**Practical related to gene amplification**			**CO5**
	A & B	Designing of primers for CDs and partial sequences			
	C	Performing PCR reactions			
	Mode of examination	Practical and/or Viva			
	Weightage Distribution	CA	MTE	ETE	
		60%	0%	40%	
	Textbook/s	Michael, R. G., Sambrook. J., "Molecular Cloning-A Laboratory Manual", 4th edition, Cold Spring Harbor Laboratory Press, 2012.			
	Other References	1. Davis, L. (2012). Basic methods in molecular biology. Elsevier. 2. Chard, T., Work, T. S., & Work, E. (1987). Laboratory techniques in biochemistry and molecular biology. Elsevier, Amsterdam.			

MMB159 : Enzymology Lab

L-T-P 0-0-3

School: SBSR		**Batch: 2020-22**			
Program: M.Sc.		**Current Academic Year: 2020-21**			
Branch: Microbiology		**Semester: 01**			
1	Course Code				
2	Course Title	**Enzymology Lab**			
3	Credits	2			
4	Contact Hours (L-T-P)	0-0-3			
	Course Status	**Compulsory**			
5	Course Objective	To give students a thorough understanding of enzymes and enzyme kinetics. To make students learn the working and operation of enzymes as well as measurement of enzyme activity			
6	Course Outcomes	CO1: To understand the mode of action of salivary amylase CO2: Preparation of standard curve for calculation of enzyme activity. CO3: Assaying the activity of industrially important amylase enzyme using 3,5-Dinitrosalicylic acid method. CO4: To determine the pH optima of amylase enzyme CO5: To determine the temperature optima of amylase enzyme			
7	Course Description	This course is designed to make students learn about enzymes, measurement of their activity in terms of IU and katal as well as understanding the kinetics of enzymes.			
8	Outline syllabus				CO Mapping
	Unit 1	**Salivary amylase**			CO1
		Mode of action of α-amylase on starch			CO1
	Unit 2	**Calculation of Enzyme Activity**			CO2
		Preparation of standard curve			CO2
	Unit 3	**Assaying the activity of industrially important amylase**			CO3
		3’5’- Dinitrosalicylic acid method			CO3
	Unit 4	**pH optima**			CO4
		To determine the pH optima of amylase enzyme			CO4
	Unit 5	**Temperature optima**			CO5
		To determine the temperature optima of amylase enzyme			CO5
	Mode of exam	Jury/Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		60%	0%	40%	
	Textbook/s*				

	Other References	

MSB116: Bioinstruments

L-T-P: 4-0-0 Credit - 4

School : SBSR		**Batch : 2020–22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Biotechnology		**Semester: 02**	
1	Course Code	**MSB116**	
2	Course Title	**Bioinstruments**	
3	Credits	4	
4	Contact Hours (L-T-P)	4-0-0	
5	Course Objective	Allow students to familiarize themselves with the specific requirements of biomedical instrumentation and biotechnology tools for enabling their intended use for research and industrial application.	
6	Course Outcomes	1. Perform experiments based on electrophoresis for separating proteins and nucleic acids. 2. Purify compounds from a mixture using column, ion-exchange, affinity chromatography, HPLC, affinity and gas chromatography. 3. Illustrate organelle and protein localization by microscopy. 4. Isolate cells by using fluorescence activated cell sorting (FACS) or magnetic activated cell sorting (MACS) and compare cell disruption techniques. 5. Conduct enzymatic and end-point assays using spectrophotometer, apply spectroscopy techniques to understand the structure of biological material.	
7	Outline syllabus:		
7.01	**Unit 1**	**Electrophoresis**	CO1
7.02	Unit 1a	Principle of electrophoresis	
7.03	Unit 1b	Agarose gel and 2D-gel electrophoresis: Principle and applications,	
7.04	Unit 1c	Capillary and Immunoelectrophoresis: Principle and applications	
7.05	**Unit 2**	**Chromatography**	CO2
7.06	Unit 2a	Paper Chromatography, TLC	
7.07	Unit 2b	Column chromatography. Ion-exchange and Affinity chromatography	
7.08	Unit 2c	Instrumentation and applications HPLC: Instrument setup and working	

7.09	**Unit 3**	**Microscopy**	CO3
7.10	Unit 3a	Principle of microscope, Optical microscopy	
7.11	Unit 3b	AFM and Fluorescence Microscopy,	
7.12	Unit 3c	Electron Microscopy	
7.13	**Unit 4**	**Cell Separation Techniques and Centrifugation**	CO4
7.14	Unit 4a	Cell isolation and cell disruption techniques	
7.15	Unit 4b	FACS and MACS- Principle and applications; Preparative centrifugation	
7.16	Unit 4c	Differential and density gradient centrifugation, Ultracentrifugation	
7.17	**Unit 5**	**Spectrometry and Spectroscopy**	CO5
7.18	Unit 5a	Spectroscopy- Absorption and fluorescence, Atomic and Raman spectroscopy	
7.19	Unit 5b	Mass spectrometry and NMR: Instrumentation and working	
7.20	Unit 5c	X-ray crystallography: crystal preparation, working and uses.	
8	Course Evaluation		
8.1	Course work: 30 marks		
8.2	Attendance	None	
8.3	Quizzes	Three best quizzes out of Five 30-minutes quizzes in lecture hours; 10 percent	
8.4	Presentations	One: 10 percent	
8.5	Assignments	Three best out of five; 10 percent	
8.6	MST	One; 20 percent	
8.7	End-term examination: 50 percent		
9	References		
9.1	Textbook	1. Wilson K. and Walker J., “Principles and Techniques of Biochemistry and Molecular Biology”, Cambridge University Press, 2010.	
9.2	Other references	1. Ninfa A.J., Ballou D.P. and Benore M., “Fundamental Laboratory Approaches for Biochemistry and Biotechnology”, Wiley, 2009. 2. Sheehan D., “Physical Biochemistry: Principles and Applications”, Wiley, 2009	

MMB110: Mycology, Phycology and Virology

L-T-P: 4-0-0 Credit - 4

School: SBSR		**Batch : 2020 – 22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 02**	
1	Course Code	MMB110	
2	Course Title	**Mycology, Phycology and Virology**	
3	Credits	4	
4	Contact Hours (L-T-P)	**4-0-0**	
	Course Status	Compulsory	
5	Course Objective	1. To prepare students with a basic understanding of algal, fungal and viral characteristics 2. To help the students understand the vegetative, asexual and sexual stages of life cycles in algae and fungi as well as lytic and lysogenic mode of reproduction in virus. 3. To impart knowledge to students about different types of viruses and their applications 4. To explain the economic importance of algae and fungi in the ecosystem	
6	Course Outcomes	After successfully completion of this course students will be able to: CO1: To understand the basics of phycology, mycology and virology CO2: To understand the mechanism of reproduction in algae, fungi and viruses CO3: Describe the life cycle of Animal Viruses, Bacterial and Plant Viruses CO4: Detailed overview on Modes of diagnosis of viruses and their applications CO5: Economic Importance of Algae and fungi	
7	Course Description	The course gives an insight into the morphology, physiology and mode of reproduction of selected algae, fungi and viruses as well as their role in the environment, agriculture, biotechnology, industry and disease. It provides a foundation for careers in microbiology, food industry, environment and biotechnology.	
8	Outline syllabus		CO Mapping
	Unit 1	**Unit I: Introduction to Phycology, Mycology and virology**	CO1
	A	Introduction to Phycology: Occurrence and distribution, physiology, pigment systems	
	B	Introduction to Mycology: Occurrence and distribution, General characteristics, Nutrition	
	C	Introduction to Viruses: Properties of virus: morphology and ultra-structure; Classification and nomenclature of viruses; Concept of viroids, virusoids, and prions	
	Unit 2	**Reproduction in algae, fungi and viruses**	CO2

	A	Reproduction in algae: Reproduction and life cycle of any three representative species from the various classes of algae			
	B	Reproduction in fungi: Generalized life cycle of any three representative members from various classes of fungi			
	C	Replication strategies in viruses: Concept of early and late proteins; Mode of transmission in plants and animals; Cell to cell transmission, Persistent and non-persistent mode of transmission			
	Unit 3	**Animal Viruses, Bacterial and Plant Viruses**			CO3
	A	Animal Viruses: Lifecycle of DNA viruses (Adenoviruses, Parvoviruses); Lifecycle of RNA viruses (Paramyxo, Toga, Rota); Lifecycle of retroviruses; Oncogenic viruse, Viral vaccines.			
	B	Bacterial: Bacteriophages: lifecycle- Lytic and Lysogenic;			
	C	Plant Viruses: Lifecycle of DNA viruses (Geminivirus); Lifecycle of RNA virus (Tobacco mosaic virus).			
	Unit 4	**Modes of diagnosis of viruses and their applications**			CO4
	A	Methods of assay: Microscopy, Histopathological changes			
	B	Infectivity assay (plague method, end point method); Serology based assay; Nucleic acid-based assay.			
	C	Application of viral vectors in cloning and expression.			
	Unit 5	**Economic Importance of Algae and fungi**			CO5
	A	Algae as pollution indicators, eutrophication agent and role in bioremediation; Role of algae in global warming and environmental sustainability			
	B	Role of cyanobacteria and selected microalgae in agriculture-biofertilizer; Production of algal pigments, biofuels and hydrogen.			
	C	Economic Importance of Fungi: Mycorrhiza: ecto-, endo-, ectendo-VAM; Fungi as insect symbionts, fungi as biocontrol agents; Potential application in Agriculture, environment, industry; Role of fungi in Biodeterioration of wood, paper, textile; Myxotoxins,			
	Mode of examination	Theory/Jury/Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Textbook/s*	1. Alexopoulos, C.J. and C.W. Mims 1979. Introduction to Mycology (3rd Ed.) Wiley Eastern Ltd N.Delhi 2. Lee, R.E. 2008. Phycology, Fourth Edition, Cambridge University Press, USA.			
	Other References	3. Kumar, H.D. 1999. Introductory Phycology. Aff. East-west Press Pvt ltd., Delhi. 4. Webster, J. and Weber, R. 2007 Introduction to Fungi. 3rd edition, Cambridge University Press, Cambridge.			

		5. Carter J. and Saunders V., (2007) "Virology: Principles and Applications", Wiley	

MMB107: Bacteriology

L-T-P: 4-0-0 Credit - 4

School: SBSR		**Batch: 2020 – 22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 02**	
1	Course Code	**MMB107**	
2	Course Title	**BACTERIOLOGY**	
3	Credits	4	
4	Contact Hours (L-T-P)	4-0-0	
	Course Status	**Compulsory** /Elective/Open Elective	
5	Course Objective	1. Understand the bacterial size, shape, arrangement and internal & external structures, bacterial growth under optimized conditions and its qualitative/ quantitate analysis, 2. Describe different modes of reproduction found in bacteria, genetic changes occurring in bacteria as evolutionary mechanisms, 3. Identify the applications of beneficial bacteria and control overgrowth of harmful bacteria.	
6	Course Outcomes	CO1: Identify bacteria on the basis of size, shape, arrangement and internal & external structures. CO2: Demonstrate bacterial growth under optimized conditions and analyse it on qualitative & quantitate parameters. CO3: Understand different modes of reproduction found in bacteria. CO4: Examine the different possibilities of genetic changes occurred in bacteria as evolutionary mechanisms. CO5: Analyse the application of beneficial bacteria and control overgrowth of harmful bacteria. CO6: Understand different aspects of bacterial life systems and relate these aspects with importance in live practices.	
7	Course Description	This course will cover the major topics in bacteriology, including bacterial size, shape, arrangement and internal & external structures, bacterial growth under optimized conditions and its qualitative/ quantitate analysis, different modes of reproduction found in bacteria, genetic changes occurring in bacteria as evolutionary mechanisms, the application of beneficial bacteria and control over growth of harmful bacteria.	
8	Outline syllabus		CO Mapping
	Unit 1	**Morphology and Fine structure of Bacteria**	CO1, CO6
	A	Size, shape and arrangement of bacterial cells	

	B	Structures external to the bacterial cell wall; cell wall composition of Gram Positive and Gram-Negative Bacteria			
	C	Other organelles internal to cell wall; spore and cysts.			
	Unit 2	**Growth and Nutrition of Bacteria**			CO2, CO6
	A	Normal growth cycle (growth curve) of Bacteria; Factors responsible for bacterial growth, synchronous growth; Continuous culture, Chemostat.			
	B	Quantitative measurement of bacterial growth (direct microscopic, plate count method); Method of isolating pure culture, pour plate and spread plate technique			
	C	Nutritional requirements and types of bacteria.			
	Unit 3	**Reproduction**			CO3, CO6
	A	Bacterial reproduction-asexual and sexual			
	B	Modes of cell division; Binary fission; Budding, fragmentation			
	C	Formation of conidiophores; septum formation.			
	Unit 4	**Bacterial Genetics**			CO4, CO6
	A	Phenotypic changes due to environmental Alterations; Genotypic changes; Mutation Types; Bacterial Recombination; Conjugation			
	B	Molecular mechanism of gene transfer by conjugation; Hfr strains, mapping bacterial genomes using Hfr strains; Transduction; Bacterial Transformation, Natural transformation and competence			
	C	Ti plasmid transfer system and its application in creating transgenics.			
	Unit 5	**Hypersensitivity and Autoimmunity**			CO5, CO6
	A	Microbes and Human welfare (medical, chemical and food industry)			
	B	Physical and chemical methods of control of Bacteria			
	C	Mode of action of Anti-microbial agents, factors responsible for controlling microbes, Physical and chemical agents.			
	Mode of examination	**Theory**/Jury/Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Textbook/s*	*Pelezar*, M.J. Reid, R.D. and E.C.S. Chan, (1986*) Microbiology* - Tata Mc Graw Hill, New Delhi.			

	Other References	Mackie and McCartney (1996) Medical Microbiology, Churchill Livingstone	

MMB108: Recombinant DNA Technology

L-T-P: 4-0-0 Credit - 4

School : SBSR		**Batch : 2020 – 22**
Program: M.Sc.		**Current Academic Year: 2020-21**
Branch: Microbiology		**Semester: 02**
1	Course Code	**MMB108**
2	Course Title	**Recombinant DNA Technology**
3	Credits	4
4	Contact Hours (L-T-P)	4-0-0
	Course Status	Compulsory
5	Course Objective	1. To illustrate creative use of modern tools and techniques for manipulation and analysis of genomic sequences. 2. To train students in strategizing research methodologies employing genetic engineering techniques.
6	Course Outcomes	After successfully completion of this course students will be able to: CO1: Recognize the ability of restriction endonucleases and other modification enzymes for genetic engineering. CO2: Apply different types of cloning and expression vectors for genetic transformation. CO3: Categorize libraries for gene isolation and use different strategies for transformation of DNA. CO4: Reframe and screen constructed libraries for differentiating between transformants and non-transformants for estimating molecular changes. CO5: Perform gene amplification using polymerase chain reaction, demonstrate DNA sequencing methods and analyse the expression of gene using RAPD, RFLP, microarray and blotting techniques. CO6: Create and formulate experiments for integrating RDT techniques for analysing manipulations and expression.
7	Course Description	The aim of this core-course is to acquaint the students to versatile tools and techniques employed in genetic engineering and recombinant DNA technology. A sound knowledge on methodological repertoire allows students to innovatively apply these in basic and applied fields of biological research. This course provides theoretical bases to properties and applications of versatile DNA modifying enzymes, cloning strategies, vector types, host genotype specificities for selection and screening of recombinants and/or recombinant transformants. Students will also be introduced to prominent nucleic acid labeling techniques. Introduction to various types of vectors viz. cloning, transformation, expression; and also vectors for genomic and cDNA library and whole genome sequencing will be provided. A critical appraisal of methods for sequencing of cloned

		genomic fragments will also be covered. This course may be deemed as a platform for introduction of more advanced cutting-edge technologies that essentially are an amalgamation of basic techniques combined in diverse forms.			
8	Outline syllabus				CO Mapping
	Unit 1	**Enzymes in r-DNA Technology**			**CO1, CO6**
	A	Introduction to gene cloning, Restriction endonucleases, ligases, alkaline phosophatase			CO1
	B	Polynucleotide kinase, terminal deoxynucleotidyl transferase, S1 nuclease, DNA polymerase I Holoenzyme, DNA polymerase III, Klenow fragment			CO1
	C	Taq DNA polymerase, RNases, ribonuclease, reverse transcriptase, poly (A) polymerase, deoxyribonuclease			CO1, CO6
	Unit 2	**Vectors for Gene Cloning and Expression**			**CO2, CO6**
	A	Essential requirements of cloning vector, Plasmids, Isolation of plasmid DNA; criteria for plasmid cloning			CO2
	B	Cloning vectors based on bacterial plasmids, bacteriophage vector for *E. Coli*, lambda replacement and insertion vectors, M13 bacteriophage			CO2
	C	Phagemids and cosmid vectors and their use vector for plant cells-Ti Plasmid; shuttle vectors; expression vectors			CO2, CO6
	Unit 3	**DNA Libraries**			**CO3, CO6**
	A	Generation of sticky and blunt ends for cloning, Linkers and adaptors, construction of genomic library			CO3
	B	construction of cDNA libraries; probe construction and labelling			CO3
	C	Methods for gene transfer-electroporation, gene gun, microinjection, liposome mediated, heat shock			CO3, CO6
	Unit 4	**Screening and Selection**			**CO4**
	A	Methods of selection and screening of recombinant DNA			CO4
	B	Introduction to antisense technology, Molecular mechanism of anti-sense technology			CO4
	C	Application of anti-sensing technology; Ribozymes and their significance in cloning			CO4, CO6
	Unit 5	**Techniques in Genetic Engineering**			**CO5, CO6**
	A	Different types of blotting techniques-Southern, northern and western			CO5
	B	RAPD, RFLP, micro array			CO5
	C	Nucleic acid sequencing (Maxam-Gilbert method and Sanger's method), Polymerase Chain Reaction and its applications			CO5, CO6
	Mode of examination	Theory			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	

	Textbook/s*	S. B. Primrose (1994). Molecular Biotechnology (2nd Edn.), Blackwell Scientific Publishers, Oxford.	
	Other References	1. J. A. Davies and W. S. Roznikolf (1992) Milestones in Biotechnology. Classic papers on genetic Engineering, Butterworth-Helnemann, Boston. 2. S. M. Kingsman and A. J. Kingsman (1998) Genetic Engineering. An Introduction to gene analysis and exploitation in eukaryotes, Blackwell Scientific Publications.Oxford. 3. Bernard R. Glick, Jack J. Pasternak, Cheryl L. Patten (2010) Molecular Biotechnology Principles and Applications of Recombinant DNA, American Society for Microbiology.	

MSB125: BIOINFORMATICS

L-T-P: 4-0-0 Credit – 4

School: SBSR	**Batch: 2020-22**			
Program: M.Sc.	**Current Academic Year: 2020-21**			
Branch: Microbiology	**Semester: 02**			
1	Course Code			
2	Course Title	**Bioinformatics**		
3	Credits	4		
4	Contact Hours (L-T-P)	4-0-0		
5	Course Objective	To acquire an advanced knowledge of bioinformatics tools used for designing and analyzing *in silico* experiments and different techniques used for molecular modeling.		
6	Course Outcomes	After successfully completion of this course students will be able to: **CO1:** Understand about overview of bioinformatics scope and their disciplines. Generation of large-scale data in the field of molecular biology. **CO2:** Review of database source, database management system, Biological databases and their classification. Sequences databases and specialized databases. **CO3:** To attain knowledge about data storage model/format, retrieval of information and integration. **CO4:** Understanding about different sequence formats. Perform sequence alignment and phylogenetic prediction with different tools/software with algorithm. **CO5:** To apply different techniques for gene prediction, motif search and genome sequencing analysis. **CO6:** Basic knowledge of various bioinformatics concepts, scope, database usage, tools and software used for each application along with their algorithms.		
7	Outline syllabus:			CO Mapping
7.01	**Unit A**		**Introduction to Bioinformatics**	CO1, CO6
7.02	Unit A Topic 1		Scope and importance	
7.03	Unit A Topic 2		Large scale generation of molecular biology data	
7.04	Unit A Topic 3		Different fields in bioinformatics	

7.05	**Unit B**		**Biological Databases**	
7.06	Unit B Topic 1		Introduction of Biological Databases	CO2, CO6
7.07	Unit B Topic 2		Structural and Sequence database	
7.08	Unit B Topic 3		Specialized Genome databases and Structure databases	
7.09	**Unit C**		**Data Storage and retrieval**	
7.10	Unit C Topic 1		Controlled vocabulary	CO3, CO6
7.11	Unit C Topic 2		Introduction to Metadata; File Storage, File Format (FASTA, GenBank, Swiss-Prot, DDBJ and PDB)	
7.12	Unit C Topic 3		Boolean Search and Fuzzy Search	
7.13	**Unit D**		**Sequence-alignment Related Problems**	
7.14	Unit D Topic 1		Sequence databases, Similarity matrices, pairwise alignment and BLAST	CO4, CO6
7.15	Unit D Topic 2		Sequence assembly and multiple sequence alignment	
7.16	Unit D Topic 3		Clustal and phylogenetics, distance based approaches, parsimony	
7.17	**Unit E**		**Sequence pattern analysis & System-wide Analysis**	
7.18	Unit E Topic 1		Structure of Prokaryotic and Eukaryotic gene, Basic and advanced sequencing (Maxam–Gilbert sequencing, Sanger sequencing, NGS, Pyrosequencing)	CO5, CO6
7.19	Unit E Topic 2		Gene finding, composition-based finding, sequence motif-based finding	
7.20	Unit E Topic 3		Pattern Matching, Regular expression, Transcriptomics, Microarray technology and expression profiles	
8	Course Evaluation			
8.1	Course work: 30%marks			
8.11	Attendance	None		
8.12	Homework	Three best out of 4 assignments: 20 marks		
8.13	Quizzes	Two 30-minutes surprise quizzes in lecture hours: 10 marks		
8.14	Projects	None		
8.15	Presentations	None		
8.16	Any other	None		
8.2	MTE	One, 20 percent		
8.3	End-term examination: 50 percent			
9	References			

9.1	Text book	Jin X., “Essential Bioinformatics”, Cambridge University Press, 2006.
9.2	Other References	1. Mount D.W., “Bioinformatics: Sequence and Genome Analysis”, Cold Spring Harbor Laboratory Press, 2004. 2. Baxevanis A., Ouellette F.B.F., “Bioinformatics: A practical guide to the analysis of genes and proteins”, Wiley-Interscience, 2004. 3. Bourne P.E., Gu J., “Structural Bioinformatics”, Wiley-Blackwell, 2009.

MMB156: Recombinant DNA Technology Lab

L-T-P: 0-0-3 Credit - 2

School: SBSR		**Batch: 2020 – 22**
Program: M.Sc.		**Current Academic Year: 2020-21**
Branch: Microbiology		**Semester: 2**
1	Course Code	**MMB156**
2	Course Title	**Recombinant DNA Technology Lab**
3	Credits	2
4	Contact Hours (L-T-P)	0-0-3
	Course Status	Compulsory
5	Course Objective	1. To illustrate creative utility of modern tools and techniques for manipulation of genomic sequences. 2. To expose students to application of recombinant DNA technology in biotechnological research. 3. To train students in strategizing research methodologies employing genetic engineering techniques. 4. To acquaint the students for analysing modification carried out in genomic sequences.
6	Course Outcomes	CO1: Development of an ability to design and conduct genetic engineering experiments. CO2: Development of an ability to analyse and interpret data of modified genomic/proteomic nature. CO3: Amalgamation of tools for creating diversification in genome. CO4: Perform time course analysis of gene expression CO5: Development of research aptitude and technical skills to secure a job in genetic engineering.
7	Course Description	The aim of this course is to acquaint the students about versatile tools and techniques employed in genetic engineering. A sound knowledge on methodological repertoire allows students to innovatively apply these in basic and applied fields of biological research. This course provides applied part of the theory by utilizing DNA modifying enzymes, cloning strategies, vector types, host genotype specificities for selection and screening of recombinants and/or recombinant transformants. This course may be deemed as a foundation course serving as a platform for introduction of

		more advanced cutting-edge technologies that essentially are an amalgamation of basic techniques combined in diverse forms and sequence.	
8	Outline syllabus		CO Mapping
	Unit 1	**Practical based on introduction to genetic engineering lab**	**CO1**
	A	Aseptic conditions maintenance in laboratory	
	B & C	Conditions optimization for growth of bacterial culture for experiments	
	Unit 2	**Practical related to preparation of plasmids and transformations**	**CO2**
	A	Plasmid isolation	
	B	Preparation of competent cells	
	C	Transformation of plasmid into competent cells	
	Unit 3	**Practical related generation of cohesive and blunt ends**	**CO3**
	A & B	Restriction of plasmid with cohesive end and blunt cutters a	
	C	Analysing the generated blunt/sticky ends using electrophoresis	
	Unit 4	**Practical related to in silico analysis of genome**	**CO4**
	A	Sequence similarity search with freely available tools	
	B	Construction of phylogenetic tree	
	C	Identification of motifs and domain in sequences	
	Unit 5	**Practical related to PCR**	**CO5**
	A & B	Designing of primers for CDs and partial sequences	
	C	Performing PCR reactions	
	Mode of examination	Practical and/or Viva	
	Weightage Distribution	CA: 60% / MTE: 0% / ETE: 40%	
	Textbook/s	Michael, R. G., Sambrook. J., "Molecular Cloning-A Laboratory Manual", 4th edition, Cold Spring Harbor Laboratory Press, 2012.	
	Other References	Frederick. M., Ausubel., Brent R., Kingston. R. E., Moore D.D., Seidman J. G., John A. Smith and Kevin Struhl, "Current Protocols in Molecular Biology", John Wiley& Son, Inc., 2003.	

MSB160: Bio-Instrumentation Lab

L-T-P: 0-0-3 Credit – 2

School: SBSR		**Batch: 2020-22**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 02**	
1	Course Code	**MSB160**	
2	Course Title	**Bio-Instrumentation Lab**	
3	Credits	2	
4	Contact Hours (L-T-P)	0-0-3	
	Course Status	**Compulsory**/Elective	
5	Course Objective	To give students a thorough understanding of tools and techniques in Biomedical and Biotechnology Laboratories. To make students learn the working and operation of various biotechnological instruments	
6	Course Outcomes	CO1: Operate autoclave, Laminar Air flow and Hot air oven and sterilize glass and plasticwares. CO2: Operate centrifuge and refrigerated centrifuge and separate cell components. CO3: Separate and visualize nucleic acids and proteins using gel electrophoresis. CO4: Operate spectrophotometer and perform absorbance assays. CO5: Separation of pigments, drugs, amino acids and hormones using chromatographic techniques. CO6 : Operation and working of different instruments and bioanalytical techniques	
7	Course Description	This course is designed to make students learn about various instruments and techniques of biomedical and biotechnology laboratory and will also enable them to use and apply these techniques and equipments to solve experimental problems.	
8	Outline syllabus		CO Mapping
	Unit 1	**Practical based on Sterilization**	CO1
		To learn the working of an autoclave.	CO1
		To learn the working of a laminar air flow.	
		To sterilize glasswares using hot air oven.	
	Unit 2	**Practical related to centrifuge**	CO2
		Using pH meter	CO2
		Working and principle of incubator shaker	
		Working of refrigerated centrifuges	
	Unit 3	**Practical related to gel electrophoresis**	CO3
		Separation of DNA using AGE	CO3

		Separation of proteins using PAGE			
	Unit 4	**Practical related to spectrophotometer**			CO4
		Principle and working of a spectrophotometer			CO4
		Measuring concentration of protein using spectrophotometer			
	Unit 5	**Practical related to chromatography**			CO5
		Use of paper chromatography for separation of plant pigments			CO5
	Mode of exam	Jury/Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		60%	0%	40%	
	Textbook/s*	Wilson K. and Walker J., "Principles and Techniques of Biochemistry and Molecular Biology", Cambridge Press, 2010.			
	Other References	1. Cottenil R.M.S., "Biophysics: An Introduction", John Wiley and Sons, 2002. 2. Gupta A., "Instrumentation and Bioanalytical Techniques", Pragati Prakashan, 2009.			

MMB160: Mycology, Phycology and Virology Lab

L-T-P: 0-0-3 Credit – 2

School: SBSR		**Batch: 2020-2022**	
Program: M.Sc		**Current Academic Year: 2020-2021**	
Branch: Microbiology		**Semester: 2**	
1	Course Code	**MMB160**	
2	Course Title	**Mycology, Phycology and Virology Lab**	
3	Credits	2	
4	Contact Hours (L-T-P)	0-0-3	
	Course Status	Compulsory	
5	Course Objective	• To train the students in microscopy of thallus structure of fungi and algae • To develop understanding of reproductive structures of fungi and algae • To learn about stages of cellular processes and cell cycle • To understand economic importance of algae and fungi • To develop knowledge of various viruses infecting plants • To give students a thorough understanding of various techniques to detect viruses in infected plant tissues	
6	Course Outcomes	CO1: Understand the morphological characteristics of algae and fungi under microscope CO2: Recollect the methods of algal and fungal culture and extraction of bioactives CO3: Appreciate the industrial and social importance of fungi and algae CO4: Understand safety measures in Virology laboratory CO5: Understanding of various techniques to detect viruses in infected plant tissues CO6: Learn mechanical dissemination of plant viruses	
7	Course Description	The course gives an insight into the morphology and physiology of selected algae and fungi, their role in the environment, agriculture, biotechnology, industry and disease. It provides a practical foundation for careers in microbiology, food industry, environment and biotechnology. It also imparts knowledge of various viruses infecting plants and a thorough understanding of various techniques to detect viruses in infected plant tissues.	
8	Outline syllabus		CO Mapping
	Unit 1	**Experiment related to fungal characteristics**	CO1
		To examine bread mould under the microscope	
		To compare morphological features (microscopic) of different classes of fungi	

	Unit 2	**Experiment related to algal characteristics**			CO1
		To compare morphological features (microscopic) of different classes of algae			
	Unit 3	**Experiment explaining viral characteristics**			CO4
		Safety measures in virology lab Detecting virus antigens through ELISA/dot blots			
	Unit 4	**Experiment demonstrating virus infecting plants**			CO5, CO6
		Identification of various virus infected plant tissues			
		PCR to detect DNA of banana bunchy top DNA virus			
	Unit 5	**Experiment demonstrating economically important fungi and algae**			CO2, CO3
		To examine edible mushroom under the microscope			
		To inspect aquatic algae/extract economically important pigment from algae			
	Mode of examination	Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		60%	0%	40%	
	Text book/s*	1. Lee, R.E. 2008. Phycology, Fourth Edition, Cambridge University Press, USA. 2. The Elements of Plant Virology- Basic Concepts and Practical Class Exercises by S.J. Kolte and A.K. Tewari			
	Other References	Lab manual			

LIST OF EXPERIMENTS

1. To examine bread mould under the microscope
2. To examine edible mushroom under the microscope
3. To compare morphological features (microscopic) of different classes of fungi
4. To compare morphological features (microscopic) of different classes of algae
5. To inspect aquatic algae/extract economically important pigment from algae
6. Safety measures in virology lab
7. Identification of various virus infected plant tissues
8. Detecting virus antigens through ELISA/dot blots
9. PCR to detect DNA of banana bunchy top DNA virus

MMB201: Environmental Microbiology & Waste Management

L-T-P: 4-0-0 **Credit: 4**

School: SBSR		**Batch: 2019 – 21**
Program: M.Sc.		**Current Academic Year: 2020-21**
Branch: Microbiology		**Semester: 3**
1	Course Code	**MMB201**
2	Course Title	**Environmental Microbiology & Waste Management**
3	Credits	4
4	Contact Hours (L-T-P)	4-0-0
	Course Status	Compulsory
5	Course Objective	1. This course provides a comprehensive introduction to microbial ecology and fundamentals of microbial diversity. 2. The course is designed to give students an up-to-date understanding of a wide array of applications of microorganisms in maintaining biogeochemical factors. 3. This course also focuses on concepts of applied environmental microbiology and how microbes can be used for various industrial/ research applications. 4. The course also highlights the modern methods of waste management and significant role of microorganisms in waste and resources management.
6	Course Outcomes	After the successful completion of this course students will be able to: CO1: Comprehend ecological interactions and role of microorganisms played in there and discuss microbial ecology concepts including methods of assessing microbial diversity and studying microbial populations. CO2: Analyze the role of microorganisms in biogeochemical cycles. CO3: Classify different methods of bioremediation and use of microorganisms and plasmids in bioremediation CO4: Explain the commercial application of microorganisms in extraction of metals, oil and in production of biogas. CO5: Identify different methods of waste management and how different microbial metabolic processes can assist in waste management.
7	Course Description	The 'Environmental Microbiology and Waste Management' is a course designed to give students knowledge about basic concepts of environment/ ecosystem and the role microorganisms play in maintaining the ecosystem balance. This course throws light on various unconventional uses of microorganisms in various industries and environmental benefits of use of the microorganisms. This course

		also outlines various biological methods of waste management and application of microbes in bioremediation.			
8	Outline syllabus				CO Mapping
®t r	**Unit 1**	**Microbial Ecology**			
	A	Ecological Concepts: Introduction to ecosystem; types of ecosystem; food chain and food web; biological magnification and eutrophication			CO1
	B	Microbial diversity: estimates of total number of species; Shannon and Simpsons indices of microbial diversity, Unculturable bacteria			
	C	Culture independent molecular methods for understanding microbial community- Partial and whole community analysis			
	Unit 2	**Role of Microorganisms in Environment**			
	A	Role of microbes in biogeochemical cycles: nitrogen cycle: different phases of nitrogen cycle, microbes involved in different stages of nitrogen cycle			CO2
	B	Carbon, Phosphorous and Sulphur cycle			
	C	Production of microbial bio-fertilizers, bio-pesticides, soil conditioners to enhance crop yields.			
	Unit 3	**Role of Microorganisms in Remediation**			
	A	Bioremediation- *in situ* and *ex situ* techniques			CO3
	B	Biodegradation of recalcitrant compounds-lignin, pesticides; Bioaccumulation of metal and detoxification			
	C	Degradation of xenobiotics by microorganisms; Degradative plasmids			
	Unit 4	**Role of Microorganisms in Mining and Energy Production**			
	A	Microbial technology in mining: Bioleaching; Biomining; Bio-beneficiation			CO4
	B	Recovery of oil and MEOR; Bioconversions			
	C	Microbial technology for energy production- Concept of microbial fuel cell- principle; types and applications, Use of microorganisms in the production of biogas			
	Unit 5	**Role of Microorganisms in Waste Management**			
	A	Landfill- structure and types, involvement of microbes in initial adjustment phase, transition phase, acid phase			CO5
	B	Methane formation and maturation phase of a landfill operation			
	C	Compositing- types; Design and operational consideration of microbial composting			
	Mode of examination	Theory			
		CA	MTE	ETE	

	Weightage Distribution	30%	20%	50%	
	Text book/s*	**1. Environmental Science.** Ahluwalia VK, Malhotra S. Ane Books India @2006. ISBN 81-8052-023-4. **2. Environmental science.** Miller GT, SpoolMan ES. 14th Edition. Brooks/Cole @2013. ISBN 13: 978-81-315-2473-2.			
	Other References	**1. Environmental Biotechnology.** Fulekar MH. CRC Press @2014. ISBN 978-1-57808-528-8. **2. Fundamentals of Ecology.** Odum EPO and Barret W. Brooks/Cole @2005. ISBN 0534420664.			

MMB202: Infection, Immunity and Diagnostics

L-T-P: 4-0-0 Credit - 4

School: SBSR		**Batch: 2019 – 21**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 03**	
1	Course Code	**MMB202**	
2	Course Title	**Infection, Immunity and Diagnostics**	
3	Credits	4	
4	Contact Hours (L-T-P)	4-0-0	
	Course Status	**Compulsory**/Elective/Open Elective	
5	Course Objective	1. Understand the infection and cells and organs of the immune system 2. Understand cell receptors and immune responses. 3. Understand the structure and function of antigens and antibodies, Ag-Ab reactions and Diagnostic Methods	
6	Course Outcomes	CO1: To understand infectious diseases, host-parasite relationship and Immunity and its types against infectious agents; To understand immune response and complement system CO2: To understand the process haematopoiesis and maturation of cells and organs of the immune system. CO3: To understand the role of various cell receptors and activation of B and T lymphocytes; cell mediated cytotoxicity and Hypersensitivity. CO4: the structure and functions of antigen and antibodies; Hybridoma technology and vaccines CO5: To understand the Antigen-Antibody Reactions and Diagnostic Methods	
7	Course Description	The objectives for the course are to acquire a fundamental working knowledge of the basic principles of immunology; to begin to understand how these principles apply to the process of immune function; and to develop the ability to solve problems in clinical immunology by making use of existing tools and techniques	
8	Outline syllabus		CO Mapping
	Unit 1	**Infection and Immune System**	
	A	Introduction to infectious diseases, host-parasite relationship, epidemiology, Immunity to infectious agents Bacteria, inter-cellular parasites, helminthes and viruses	CO1
	B	First, second and third line of defense; Immunity-innate and acquired immunity;	CO1
	C	Cell-mediated and humoral immunity; Phagocytosis; Complement system and inflammatory responses	CO1

	Unit 2	**Cells and Organs of the Immune System**			
	A	Haematopoesis and maturation of immune cells			CO2
	B	Organ and cells of the immune system-primary and secondary lymphoid organ			CO2
	C	B-lymphocytes, T-lymphocytes, macrophages, dendritic cells, langerhan cells, Natural killer cells, eiosinophils, basophils, neutrophils and mast cells			CO2
	Unit 3	**Cell Receptors and Immune Responses**			
	A	BCR, TCR and MHC; Activation of B and T- lymphocytes; Generation of humoral cell and cell mediated immune responses			CO3
	B	Cell-mediated cytotoxicity; Antibody-dependent cell mediated cytotoxicity; Macrophage-mediated cytotoxicity;			CO3
	C	Hypersensitivity; Autoimmunity; Cytokines and their role in immune regulation.			CO3
	Unit 4	**Antigen and Antibody**			
	A	Nature, biology and types of antigens and super antigens; epitopes; adjuvants			CO4
	B	Antibody structure, types and functions; Hybridoma technology and monoclonal antibodies			CO4
	C	Vaccine and type of vaccines.			CO4
	Unit 5	**Antigen-Antibody Reactions and Diagnostic Methods**			
	A	Antigen-antibody reactions-agglutination and precipitation			CO5
	B	Immunological methods-ELISA, RIA			CO5
	C	Immunodiffusion, Immunofluorescence, complement fixation test etc.			CO5
	Mode of examination	**Theory**/Jury/Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Textbook/s*	Kindt T.J., Osborne B.A. and Goldsby R.A. (2006) Kuby Immunology, W. H. Freeman			
	Other References	1. Delves P.J, Martin S.J., Burton D.R. and Roitt I.M., (2011) Roitt's Essential Immunology, Wiley			

MSB207: Microbial Biotechnology

L-T-P: 4-0-0 Credit: 4

School : SBSR	**Batch : 2019-21**
Program: M.Sc.	**Current Academic Year: 2020-21**

Branch: Microbiology		**Semester: 3**	
1	Course Code	**MSB207**	
2	Course Title	**Microbial Biotechnology**	
3	Credits	4	
4	Contact Hours (L-T-P)	4-0-0	
5	Course Status	Compulsory	
6	Course Objective	1. Some Potential Sources of Components of Industrial Media 2. Product recovery, Solids (Insolubles) Removal 3. Industrial production of organic acids 4. Role of microorganisms in hydrocarbon degradation	
7	Course Outcomes	After studying this course, students will be able to CO1: Determine Primary and Secondary screening, Production strains, and Production media CO2: Evaluate Filtration; Centrifugation; Coagulation and flocculation CO3: Interpret the production of microbial insecticides, production of Biopolymers, Biofuels CO4: Analyze the role of microorganisms in hydrocarbon degradation CO5: Determine Role of microorganism in Bioleaching and Textile Industry. CO6 : Analyze types of microorganisms found on textile fibres	
8	Course Description	This course contains introductory part of industrial biotechnology which includes various useful microorganisms, their production, different types of fermentors, product recovery processes. After this course study student will able to learn the role of microorganisms in textile industry and marine environment.	
9	Outline syllabus		CO Mapping
	Unit 1		**CO1**
	A	Introduction and history, Isolation and screening, Primary and Secondary screening, Production strains, Production media,	
	B	Raw Materials Used in Compounding Industrial Media, Growth Factors, Water,	
	C	Some Potential Sources of Components of Industrial Media, Inoculum preparation, Introduction to Fermenter, Industrial sterilization	
	Unit 2	**Product recovery, Solids (Insolubles) Removal**	**CO2**
	A	Filtration; Centrifugation; Coagulation and flocculation;	
	B	Foam fractionation; Whole-broth treatment; Primary Product Isolation : Cell disruption;	
	C	Liquid extraction; Dissociation extraction ;Ion-exchange adsorption; precipitation	
	Unit 3		**CO3**
	A	Introduction, Industrial production of penicillin, production of streptomycin	
	B	Industrial production of organic acids- production of citric acid, lactic acid, amino acids such as L- glutamic acid,	

		production of single cell proteins, production of fermented foods,			
	C	Production of microbial insecticides, production of Biopolymers, Biofuels, Production of Alcohol Yeasts , food yeast and Baker's Yeast.			
	Unit 4	**Petroleum Microbiology**			**CO4**
	A	Types of compounds in petroleum, products of compounds in petroleum, Microorganisms in hydrocarbon system			
	B	Role of microorganisms in hydrocarbon degradation.			
	C	Marine Microbiology: Characters of marine environment, characters of marine microorganisms, role of marine microorganisms			
	Unit 5				**CO5**
	A	Production of Vaccines -Production of virus vaccines; Production of bacterial toxoids; Production of killed bacterial vaccines;			
	B	Role of microorganism in Bioleaching and Textile Industry : A. Bioleaching of elements – Microorganisms involved, chemistry of microbial leaching and beneficiationB			
	C	Textile Industry – Types of microorganisms found on textile fibres,Prevention of growth of microorganisms.			
	Mode of examination	Theory			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Text book/s*	1. Crueger & Crueger Biotechnology: A Text Book of Industrial microbiology 2nd edition 2. Demain, A.L Biology of Industrial Microorganisms			
	Other References	1. Hobbs, B.C. and Rioberts,D 1993 Food Poisoning and Food Hygiene Edward Anold, London. 2. Hui Y H 2006 Food Biochemistry and Food Processing Blackwell 5. Joshi, V.K. Ashok Pondey 1999 Biotechnology and Food fermentation Vol. I & II. 3. Patel, A.H. Industrial microbiology			

MMB207: Fermentation and Downstream Processes

L-T-P: 4-0-0 **Credit - 4**

School: SBSR		**Batch: 2019 – 21**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 3**	
1	Course Code	**MMB207**	
2	Course Title	**Fermentation and Downstream Processes**	
3	Credits	4	
4	Contact Hrs (L-T-P)	4-0-0	
	Course Status	Compulsory	
5	Course Objective	1. To enable students bridge the gap between theoretical concepts and practical aspects in fermentation technology. 2. To provide knowledge about the different processes being used to prepare various industrially important substances 3. To enable students to understand the bioreactor designs. 4. To provide insight of various downstream process.	
6	Course Outcomes	CO1: Understand the history of fermentation technology and growth kinetics of microorganisms. CO2: Design bioreactors to achieve desired results (i.e. specified cell concentration, production rates, etc). CO3: Examine the mass transfer operation of various biochemical processes. CO4: Development of industrial fermentation process. Justify the use of different biochemical strategies for the production and separation of biologicals.	
7	Course Description		
8	Outline syllabus		CO Mapping
	Unit 1		**CO1**
	A	Fermentation, basic concept, submerged and solid state fermentation	
	B	Microbial growth kinetics, Microbial nutrient requirements,	
	C	Sterilization of media, air and equipments for fermentation	
	Unit 2		
	A	Batch, Continuous and Fed batch mode of operation	**CO2, CO3**
	B	Operational design of Bioreactor- vessel, agitator, sparger, baffles, types of Bioreactors- STR, CSTR, Airlift fermenter,	
	C	Fluidized bed reactor, Packed bed reactor, Immobilized cells and enzymes bioreactor	

	Unit 3				**CO2, CO3, CO4**
	A	Measurement, monitoring and control of physical, chemical and biological parameters in a bioreactor			
	B	Transport phenomena in bioreactor			
	C	Aeration and agitation in bioreactors; pH and temperature control in bioreactor			
	Unit 4				**CO2, CO3, CO4**
	A	Cell disruption methods for intracellular products-Osmotic and heat shock, Homogenization, Sonication, Freezing thawing, Enzyme digestion.			
	B	Centrifugation: basic principles, design characteristics and applications			
	C	Membrane based separation processes,			
	Unit 5				**CO3, CO4**
	A	Chromatographic techniques			
	B	Electrophoretic separation			
	C	Evaporation, drying and crystallization techniques.			
	Mode of examination	Theory/Jury/Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Textbook/s *	1. McNeil B. and Harvey L., "Practical Fermentation Technology", Wiley, 2008.			
	Other References	2. Doran P.M., "Bioprocess Engineering Principles", Academic Press, 2012. 3. Bioseparations Principles and Techniques, B. Sivasankar. Prentice hall of India Pvt. Ltd., 2007.			

MMB208: Food Microbiology

L-T-P: 2-0-0 Credit - 2

School : SBSR		**Batch : 2019– 21**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 03**	
1	Course Code	MMB208	
2	Course Title	Food Microbiology	
3	Credits	2	
4	Contact Hours (L-T-P)	2-0-0	
	Course Status	Compulsory	
5	Course Objective	The course is designed to prepare students with a basic understanding of the microbes involved in biological processes such as fermentation and spoilage. The course provides a foundation for careers in microbiology, food microbiology, or research in all branches of food sciences.	
6	Course Outcomes	After the successful completion of this course students will be able to: CO1. Recognize and describe the characteristics of important pathogens and spoilage microorganisms in foods. CO2. Understand the role and significance of intrinsic and extrinsic factors on growth and response of microorganisms in foods. CO3. Identify ways to control microorganisms in foods. CO4. Identify the conditions under which the important pathogens and spoilage microorganisms are commonly inactivated, killed or made harmless in foods. CO5. Utilize laboratory techniques to detect, quantify, and identify microorganisms in foods. CO6.Understand the role of fermentation and preservation in food science.	
7	Course Description	The 'Food Microbiology' course outlines the basic principles of Microbiology. This course also sheds light upon fermentation and is designed to make student learn the preservation of food products. The course also further encompasses the concept of identification and quantification of microorganisms in foods.	
8	Outline syllabus		CO Mapping
	Unit 1	**History development and microbes in food**	
	A	Historical developments	CO1, CO2
	B	Important of Microorganisms in food	
	C	Factors affecting growth of microbes in food	
	Unit 2	**Spoilage of Foods**	

	A	Spoilage of meat			
	B	Spoilage of Milk and milk products			
	C	Spoilage and defects of fermented food products			CO3, CO4
	Unit 3	**Biological transformation of food**			
	A	Fermentation			
	B	Production of fermented products			
	C	Importance of fermentation			CO3, CO6
	Unit 4	**Preservation of food**			
	A	General principles of food preservation			
	B	Chemical Preservation of food			CO6
	C	Preservation of food by radiation			
	Unit 5	**Food Borne Diseases**			
	A	Bacterial and nonbacterial infection			
	B	Food borne diseases: Salmonellosis, Botulism, Listeriosis			
	C	Detection of Microbes in food			CO4,CO5, CO6
	Mode of examination	Theory			
	Weightage Distribution	CA	MTE	ETE	
		30%	20%	50%	
	Textbook/s*	1. Jay, J.M. (2008) Modern Food Microbiology (Sixth Edition). Aspen Publishers, Inc. Gaithersburg, Maryland.			
	Other References	2.Adams, M. R. and Moss, M. O. (2005) Food Microbiology (Second edition). Royal Society of Chemistry Publication, Cambridge. 3. Ray, B. (2005) Fundamental food microbiology (Third edition). CRC Press, New York, Washington D.C. 4. Frazier, W. C. and West off, D. C. (2007) Food Microbiology. Tata McGraw Hill Publishing Company Ltd. New Delhi. 5. Banwart G J. (1989). Basic Food Microbiology. AVI publication.			

MSB259: Microbial Biotechnology Lab

L-T-P 0-0-3 Credit 2

School: SBSR		**Batch:**	
Program: M. Sc.		**Current Academic Year: 2020-21**	
Branch: BT		**Semester: 3rd**	
1	Course Code	**MSB259**	
2	Course Title	**Microbial Biotechnology Lab**	
3	Credits	2	
4	Contact Hours (L-T-P)	0-0-2	
	Course Status	Compulsory/Elective	
5	Course Objective	• To develop practical knowledge of microorganism • To teach students about fermentor; other instruments and their components • To teach about microbial production of various biomolecules	
6	Course Outcomes	CO1:Practical knowledge of fermentor other instruments and their components CO2: Isolation and screening of microorganisms CO3: Practical knowledge of solid state fermentation. CO4: Able to produce different biomolecules CO5: Cradle to grave knowledge of microbial process engineering.	
7	Course Description	**Microbial Biotechnology**, is a specialization of biotechnology, It deals with the design and development of reactor and processes for the manufacturing of products such as like enzymes, acids, biopolymers etc. This lab covers the design of bioreactor and its operations.	
8	Outline syllabus		CO Mapping
	Unit 1	**Isolation and screening of microorganism**	**CO1, CO5**
		Isolation and screening of microorganism producing proteases	
		Isolation and screening of microorganism producing amylases	
	Unit 2	**Isolation and screening of microorganism**	**CO2, CO5**
		Isolation of Nitrogen fixers from soil	
		Isolation of phosphate solubilizers from soil	
	Unit 3	**Microbial Growth Kinetics**	**CO2, CO5**
		Estimation of effect of temperature on microbial growth	
		Estimation of effect of pH on microbial growth	
	Unit 4	**Microbial fermentation**	**CO4, CO5**
		Fermentative production of Wine	
		Fermentative production of Beer	
	Unit 5	**Microbial fermentation**	**CO4, CO5**

		Fermentative production of Amylase			
	Mode of examination	Practical/Viva			
	Weightage Distribution	CA	MTE	ETE	
		60%	0%	40%	
	Text book/s*	-			
	Other References				

MMB255: Immunology Lab

L-T-P 0-0-3 Credit 2

School : SBSR		**Batch : 2020-2022**	
Program: M.Sc.		**Current Academic Year: 2020-21**	
Branch: Microbiology		**Semester: 3rd**	
1	Course Code	**MMB255**	
2	Course Title	**Immunology Lab**	
3	Credits	2	
4	Contact Hours (L-T-P)	0-0-3	
5	Course Status	Compulsory	
6	Course Objective	1) This course understanding provides a strong foundation and can prompt a greater enthusiasm for and an improved understanding of the complete immune response. 2) The Work involving human samples is enticing to students with clinical interests, and further detailed protocols, and analysis guidance may be appropriate for introductory immune response.	
7	Course Outcomes	After successfully completion of this course students will be able to: CO1: understand basic laboratory techniques of blood groups CO2: estimate the haemoglobin of its own blood CO3: practical knowledge of antigen antibody interactions CO4: isolate lymphocytes for further deep analysis CO5: prepare suspension solutions of spleen and bone marrow	
8	Course Description	The aim of this course is to acquaint the students about the versatile tools and techniques employed in immunology. The course will also provide students with a hands-on understanding of how immunology can be used to discover various processes used by animals and humans for their self defence mechanism.	
9	Outline syllabus		CO Mapping
	Unit 1		**CO1**
	A	To study permanent slides of immune tissues and organs	
	B	To find the blood group of own blood	
	C	To find the Rh factor of own blood group	
	Unit 2		**CO2**
	A	To estimate the amount of Hb present in human blood	
	B	To perform Rocket immunoelectrophoresis	
	C	To perform Separation of lymphocytes	
	Unit 3		**CO3**
	A	To perform Sandwich enzyme linked immunosorbant assay	
	B	To perform DoT ELISA	
	C	To perform Haemagglutination test	
	Unit 4		**CO4**
	A	To perform Ouchlerlony's double immunodiffusion method.	
	B	To perform Radial Immunodiffusion	

	C	To perform RIA			
	Unit 5				**CO5**
	A	Preparation of single cell suspension of spleen.			
	B	Preparation of single cell suspension of bone marrow.			
	C				
	Mode of examination	Practical/or Viva			
	Weightage Distribution	CA	MTE	ETE	
		60%	0%	40%	
	Text book/s*	Kindt, T. J., Goldsby, R. A., Osborne, B. A., Kuby, J. (2006). VI Edition. Immunology. W.H. Freeman and Company.			
	Other References	Delves, P. J., Martin, S. J., Burton, D. R., Roitt, I.M. (2006). XI Edition. Roitt's Essential Immunology, Blackwell Publishing			